

International Journal of Multidisciplinary Research and Development

www.allsubjectjournal.com

ISSN Online: 2349-4182 ISSN Print: 2349-5979

Received: 06-01-2024, Accepted: 29-01-2024, Published: 21-02-2024

Volume 11, Issue 2, 2024, Page No. 107-114

Detailed mechanism of cadmium toxicity

Aditi Ghosh*, Omprakash, Poonam Devi, Sanjanna Boya, Deepak Chopra

Department of Animal Biochemistry, ICAR-National Diary Research Institute, Karnal, Haryana, India

Abstract

The innate ability to have children is called fertility. The fertility rate is known as the number of children born per couple, individual, or population. 8-12% of couples are thought to have infertility globally. A subset of 3-5% of these couples experiences unexplained infertility. Recent estimates place the number of infertile couples at 60-80 million per year worldwide, with 15-20 million of those individuals living in India alone. People are exposed to metal aerosols at work and in the environment with the highest cadmium, which builds up in the male reproductive system. The presence of Nectin-2, a junction found in ectoplasmic specialization (ES), which is responsible for the blood-testis barrier, is the cause of the accumulation of cadmium in the male reproductive system. The preceding claim that the testis is more vulnerable to cadmium is supported by data that basal ES is the target of cadmium. Erectile dysfunction, one of the major problems underlying male infertility, affects up to 10% of men. Due to its disruption of the hypothalamus-hypophysis testis axis (HHT), impairment of the androgen receptor's conformation, and methylation of numerous genes, cadmium has been classified as an endocrine disruptor. It inhibits the STAR gene, which is in charge of moving cholesterol from external to internal mitochondria, P450scc, which aids in the conversion of cholesterol to pregnenolone, and other genes. Fecal microorganisms, or gut bacteria, play a positive role in the colon rectum by encouraging the production of short-chain fatty acids, and free amino acids, and favoring the transfer of secondary bile acids from primary bile acids. Ben Eiseman performed the first fecal microbiota transplant to cure a Clostridium difficile illness. Fecal microbiota transplantation is a novel approach that involves introducing feces into the recipient's GI tract via a variety of delivery techniques, including colonoscopy, nasogastric, nasojejunal, and others. The therapeutic effects of FMT can be explained by a mechanism known as competitive niche exclusion. An excellent illustration of how FMT restores normal host metabolism and gut microbiota is the restoration of Firmicutes phylum bacteria and secondary bile acid metabolism.

Keywords: Infertility, cadmium toxicity, fecal microbiota transplantation, gut microbiota

Introduction

The harmful effects of heavy metals on male reproduction have been a hot topic in toxicology during this massive period of industrialization and urbanization. When consumed, heavy metals—a class of metals with a density greater than that of water-always result in toxicity in the body of the consumer. Heavy metals (> 5mg/kg) include substances like lead, mercury, arsenic, cadmium, and others. But in this paper, we will be focusing on cadmium heavy metal, as ingestion of cadmium has come on the rise in the past few years. Most of the reports have shown that males are exposed to heavy metals either occupationally or environmentally or both. Intake of contaminated food is a usual source of heavy metals. Contamination can occur for a variety of reasons, including improper handling of food, food processing, transit, etc. (Kidd et al., 2002). Even a few reports have surfaced indicating the presence of heavy metals like Cd and Pb in water supplies, foods like spinach and cabbage, cereals, and cosmetics, among other things. Moreover, meta-analyses have brought about the detrimental effects of environmental pollutants on male reproduction. It has been widely known by now that environmental pollutants like a small amount of Pb, Cd, Hg, etc, pave the way to the generation of ROS. Due to cadmium's inclusion in legumes, green leafy vegetables, pesticides, and insecticides, intake has recently grown. Small in size, it enters the body quickly and builds up in different organs depending on the susceptibility (Faroon et al., 2012). Compared to non-smokers, smoking causes a 4-5 fold increase in cadmium levels. Spermatozoa are highly

vulnerable to reactive oxygen species because it consists of large amounts of membrane polyunsaturated fatty acids (PUFA). As a result, it causes DNA damage, testicular apoptosis, disruption of seminiferous epithelium, and abnormal sperm function leading to male infertility (Pant et al., 2003) [28]. Due to the lack of a detoxification mechanism in the human body, these toxins build up in the male reproductive system and reduce fertility (Wan et al., 2003) Male infertility and poor-quality sperm have earned a place as global issues. Heavy metals can damage Sertoli and Leydig cells as well as hinder the creation of testosterone. It accomplishes this by meddling at the structural and gene levels. Androgen Binding Protein (ABP), cadherins, occludins, and other neuroendocrine processes are also extremely vulnerable to these heavy metals (Chung and Cheng, 2001) [3]. It is believed that 40–50% of a couple's infertility is caused by male factors. Low sperm count, slow sperm movement, and aberrant sperm morphology, which are referred to as oligospermia, asthenospermia, and teratozoospermia, respectively, are the three most important indicators of male factor infertility (Kilchevsky et al., 2012) [22]. The ratio of Firmicutes to Bacteroides is largely thought to be important for preserving gut homeostasis and safeguarding the host from the invasion of any alien bacteria (Nelson et al., 2010) [19]. Humans contain 100 trillion microbes, therefore this directly or indirectly controls every cell in the body. According to reports, the gut microbiota has a bidirectional effect on male reproduction since some of them express 17-HSD, 3-HSD, and other steroidmetabolizing enzymes (Devendran et al., 2017) [6].

Researchers are developing novel treatments to lessen the adverse effects of medications used to reduce male metal toxicity.

Effect of Cadmium

Cadmium is harmful because of its propensity to imitate divalent cations like Ca⁺² and Mg⁺², which interfere with physiological processes in the body. Cd can cause genetic instability, protein and DNA damage, and ultimately apoptosis at the cellular level. In connection with this, it generates a great deal of oxidative stress, which in turn

causes the polyunsaturated fatty acids in the membrane to degrade, which opens the door to the disorder of membrane structure (Dixon *et al.*, 1998) [10]. When it comes to morphology, Cd causes the diameter and length of seminiferous tubules to decrease. When instances of apoptosis were found in germinal cells, the fact acquired confidence. Henceforth, Cadmium has been identified as an apoptosis inducer. Some other reports have also depicted the picture of ER expansion, mitochondrial dilation, and vacuolization after being exposed to cadmium.

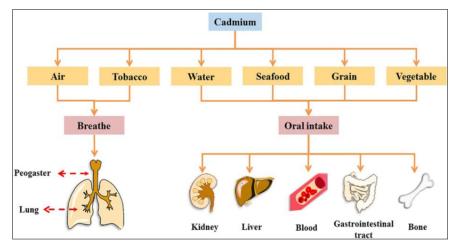


Fig 1: Major routes of Cd exposure and toxic effects of Cd on different organs in the human body (Wang et al., 2021) [45]

Numerous reports have revealed various numerical values of half-lives in various body parts. For instance, the half-life in the kidney is 6-38 years, the liver is 4-9 years, and the blood is 75–128 days, but the corporal accumulation depends on the length and dosage of exposure because it mimics the critical divalent ions required for many physiological functions (WHO 2010). It is evident from the findings above that it has a significant impact on reproduction.

Mechanism of Cadmium toxicity

Utilizing the ability to imitate the divalent ions, Cadmium can interact with membrane transporters, allowing it to enter cells. Cadmium transport proteins are members of the Ca voltage-gated channel (VGCC) or Fe Transporters family of

proteins. The digestive tract protein ZIP-8 and the divalent metal ion transporter DMT-1 have the highest Cd affinities, respectively (*Fujishiro et al.*, 2017) [16]. Male reproduction is one of the several processes that Cd mimics and interrupts. In particular, the absolute quantity of aspartate residues present at physiological pH has been demonstrated to be correlated with cadmium sensitivity and channel composition. It binds with albumin after being ingested, assisting with delivery to other organs. The liver, the body's first organ to which it gets there, later serves as the starting point for the production of metallothioneins (MT), a group of proteins with a high cysteine content that binds to metals (Fels *et al.*, 2019) [15]. Additionally, it has a strong propensity to cause oxidative

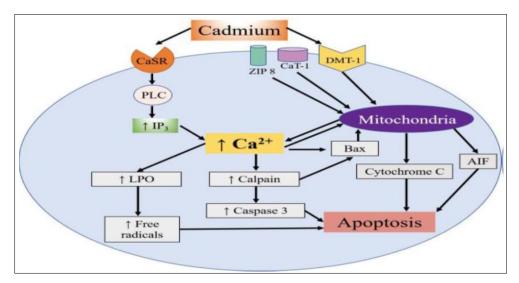


Fig 2: Apoptotic effects of cellular Cd reported in different cell types. CaSR: Ca- sensitive receptor, LPO: lipoperoxidation, DMT-1: Dimetallothionein, ZIP-8: Zinc transporter 8, AIF: Apoptosis-inducing factor (Arteaga *et al.*, 2021)

stress while subtly suppressing the action of the antioxidant enzyme. Additionally, it has extended its influence to mitochondria, where it degrades the -SH groups of cysteines found in the internal membrane proteins, uncoupling the energy that could have been retained for ATP generation. Additionally, it triggers the intrinsic process of apoptosis, which is signaled by the release of the proteins apoptosisinducing factor (AIF) and mitochondrial cytochrome C. (Thevenod et al., 2013). Additionally, it causes phospholipase C (PLC) to become activated, which raises the Ca concentration through an increase in the second inositol triphosphate messenger (IP3). By boosting the translocation of calpains (Ca-dependent cysteine proteases), these IP3 increases also cause the autolysis and activation of the pro-apoptotic protein Bax. It is further connected to the decline in GPX, CAT, and SOD activity as well as glutathione (GSH) levels. Even if Cadmium's principal target is mitochondria, it readily interacts with receptors in the plasma membrane by employing the ability to form multiple unions. This illustrates Cd's ability to modify signal transducing pathways. Another way of Cd toxicity includes alteration in the adhesion and migration since it can tune the structure of the cytoskeleton by modifying the concentrations and localizations of E-cadherins, Ncadherins, and β-catenin proteins (Prozialeck et al., 2003) [31]. These proteins altogether are important components of tissue integrity. The mechanism behind modifying the structure is the mimicking power of the Cd.

Blood Testis Barrier: The major target of Cd

The blood-testis barrier is disrupted by heavy metals, most notably Cadmium, through certain signal-transduction pathways and particular proteins. Tight junctions (TJ), desmosome-like junctions (DLJ), gap junctions, and-most significantly—basal ectoplasmic specialization together to form the blood-testis barrier. The basal ES is now an actin-based adherens junction that is exclusive to the testis. It should be highlighted that cohabitation is necessary to allow for periodic BTB reorganization to support stage VIII preleptotene spermatocyte migration. Due to its unusual morphological structure, it is susceptible to Cadmium. The principal target of the Cd is E-cadherin. Due to the calcium-binding motif on E-cadherin, Cd interacts with the motif and interferes with cell adhesion (Yan and Cheng, 2005) [48].

Signal transduction pathways control how harmful Cd is at the blood-testis barrier. When exposed to cadmium, the BTB produces more TGF-3, which in turn triggers the p38 MAPK signaling pathway. As a result, the levels of integral membrane proteins (such as occludin, N-cadherin, and E-cadherin) near the BTB location decrease. Additionally, proteases are activated, which further compromises the BTB's integrity and, as a result, results in the loss of germ cells from the seminiferous epithelium. This activates the c-JNK signaling pathway to inhibit proteolysis (Wong *et al.*, 2005) [35]. Few experiments have also reported the fact that

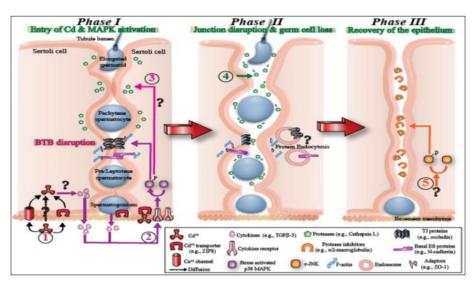


Fig 3: Cadmium-induced Blood –testis barrier (BTB) and cell junction disruption in seminiferous epithelium by activating specific signal transduction pathways. In phase 1 Cd enters the BTB barrier via Ca-gated channels or DMT in the Sertoli cells. After entering the Sertoli cells, it induces the synthesis of the cytokine TGF-β3, which interacts with their respective receptors and activates the stress-activated p38 MAPK signaling pathway. In phase II, BTB is disrupted via the degradation of integral membrane proteins as well as clathrin-mediated endocytosis of the tight junctions and basal ES proteins. In phase III, to limit the unwanted proteolysis, the c-JNK signaling pathway is activated which stimulates the release of proteases inhibitors (e.g α2-macroglobulin) to recover the seminiferous epithelium (Siu *et al.*, 2009)

Nectin-2 which is a junction molecule present at the basal and apical ectoplasmic specializations (ES) for the formation of the Blood-testis barrier, is crucial for spermatogenesis. It has been established that Nectin-2 is what makes BTB more susceptible to infection than other

tissues (Takai *et al.*, 2003) ^[40]. Since Nectin-2 is the flag protein for ES, it is thought that cadmium directly targets Nectin-2. According to the findings, Cd inhibits nectin-2 at the transcriptional and post-translational stages.

Table 1: The below table shows the effects of cadmium in testicular cells *in vivo*

In vitro model	Observation after exposure	References
Primary rat Sertoli cell culture	Low-dose: impairment of development and maintenance of TJ- permeability barrier	(Janecki et al., 1992) [20]

	High-dose: decrease in inhibin secretion and cell viability	(Janecki <i>et al.</i> , 1992) [20]
	Cell vacuolization and pyknosis; cell became round and then get detached from the substratum	(Syed et al., 1997)
Rat Sertoli cell line cultures (ASC-D19)	TJs disruption, apoptosis	(Sorenson and Brabee, 2003)
Primary rat Sertoli cells and primary rat		(Clough et al., 1990)
Leydig cell cultures	cells	(1 1 1 8 1 1 1 1)

The epididymis

The spermatozoids need to mature as well as leave the testis to fertilize an egg. By first passing via the epididymis, this maturation is accomplished. Three primary regions make up the epididymis: the caput, corpus, and cauda (Robaire et al., 2015) [33]. Epididymis weight falls in response to exposure to cadmium, and the corpus shows the greatest deposition of Cd (Elgawish et al., 2014) [12]. It is well-recognized that androgens are essential to the epididymis' ability to maintain a healthy epithelium. To ensure that proteins, glycoproteins, glycolipids, and phospholipids are properly secreted by the epithelium, which is necessary epididymis spermatozoids to develop and survive, T secretion is crucial (Robbaire et al., 2015). But when animals and the human gets exposed to Cd, it disrupts the androgen receptor, consequently decreasing their T concentrations and thereby reducing the size and weight of epididymis. Moreover, the epididymis is also responsible for conserving spermatozoid's genetic material while it transits to the fertilization site. Here, Cd causes the decompaction of chromatin by disrupting disulfide bridges, which makes spermatic DNA vulnerable to fragmentation (Olivia et al., 2006). Cd lowers the concentration of N-acetyl glucosamine, sialic acid, and fucose in the membrane of spermatozoids which gives them the capacity to cross the cervical mucosa (Robaire et al., 2015) [33]. However, when humans and animals are exposed to Cd, it affects the androgen receptor, which lowers their T levels and, as a result, decreases the size and weight of the epididymis. Additionally, the epididymis is in charge of protecting the genetic material of the spermatozoid while it travels to the site of fertilization. Here, Cd disrupts disulfide bridges to cause chromatin decompaction, leaving spermatic DNA prone to fragmentation (Olivia et al., 2006). Cd makes spermatozoids' membranes less concentrated in N-acetyl glucosamine, sialic acid, and fucose, allowing them to pass the cervical mucosa (Robaire et al., 2015) [33]. All of the aforementioned alterations can be directly attributed to the drop in androgens, which either directly or indirectly impedes all associated activities. Tyrosine phosphorylation is crucial for the development of spermatozoids' motility.

Cadmium as an endocrine disruptor

As a result of altering the steroidogenic area in Leydig cells, Cd controls the expression of the androgen receptor (AR) gene and reduces the ability of the body to produce testosterone. It decreases the amounts of steroidogenic acute regulatory protein (StAR), LH receptor, and cAMP in the testis (Henson *et al.*, 2004) [18]. Additionally, it affects the hypothalamus-pituitary-testicular axis, which affects hormone levels. By attaching to each hormone's receptor, cd suppresses hormone production through competitive tactics. It can also compete with Dihydroxy testosterone (DHT), a naturally occurring ligand of AR. The transcription potential

is decreased when Cd binds to AR and modifies its conformation (Lafuente *et al.*, 2003) [23]. Well, first of all, cAMP stimulates the production of T when cholesterol is transferred from the external to the internal mitochondrial membrane and kickstarts the production of pregnenolone with the aid of the enzyme P450scc. PKA is simultaneously activated by cAMP, and PKA in turn activates transcription factors that code for steroidogenic enzymes like StAR. Pregnenolone is now transformed into progesterone by the enzyme 3-hydroxysteroid dehydrogenase (3-HSD). 17-hydroxysteroid dehydrogenase (17-HSD) controls the conversion process, which results in androstenedione and eventually testosterone. Numerous investigations have demonstrated that Cd decreases the expression and activity of the 3-HSD and 17-HSD enzymes

Cadmium affects the immune system

The hypothalamus-hypophysis axis is damaged by Cd, which has long been known. Numerous cytokines are associated with the HHT axis to ensure its normal operation, including the proinflammatory cytokines tumor necrosis factor (TNF-), interleukin-1, and interleukin 6 (IL-6) (Diez-Pina *et al.*, 2009) [8]. TNF-, IL-1 and IL-6 levels significantly increase when the HHT axis is compromised. Additionally, it increases the levels of interleukin 10 (IL-10) and interferon-gamma (IFN), demonstrating Cd's capacity to increase inflammatory responses. Oxidative stress levels grow in tandem with the level of inflammation.

Multiple investigations have revealed elevated levels of TNF-, IL-1, IL-6, and IL-8, which were strongly linked to Cd-induced alteration of the gut flora. Clear photos of necrotic areas, short and thick villi, and many fusions were also displayed; these features were once more connected to an increase in the cytokines discussed earlier.

Cd has direct effects on gut microbiota!

Recent data suggest that exposure to heavy metals is the leading cause of the development of several metabolic illnesses, including type 2 diabetes, obesity, male infertility, and newborn infections. Heavy metals, especially Cadmium, are thought to be targets for the gut microbiota as a "superorganism." There are many HM-binding bacterial strains in the gut microbiota, including Lactobacillus, which is been shown to be beneficial for reducing heavy metal toxicity. Exposure to heavy metals and the gut microbiota composition are correlated in both directions since heavy metals can also hinder bacterial growth. By interfering with protein synthesis, cadmium limits the growth of bacteria. The majority of research has shown that Firmicutes and Proteobacteria abundance dropped after the following exposure, but the abundance of Bacteroidetes increased (Dheer et al., 2015) [7]. All of these lead to an increase in fatty acid and lipid metabolism as well as increased

intestinal permeability. Additionally, Cd affects the gut flora in a metal-specific and time-dependent manner. In comparison to gram-negative bacteria, gram-positive bacteria are more susceptible as a result (Fazeli *et al.*, 2011) ^[14]. Similarly, Lactobacillus was less sensitive to Cd toxicity than Bifidobacteria, which may be due to Lactobacillus' considerable Cd-binding abilities (Bridou *et al.*, 2011) ^[2]. Therefore, it may be inferred from the aforementioned information that Cd can alter the composition of the gut microbiota, leading to an increase in LPS and inducing inflammation in the liver. By lowering the quantities of

Akkermansia, which is in charge of both inducing TJ protein expression and maintaining the intestinal mucus layer, it can concurrently disrupt the gut barrier junctions. The overall increasing ratio of Bacteroidetes/Firmicutes and body fat is mostly due to these changes. The fact that subsequent antibiotic treatment effectively halted the buildup thus far after fecal microbiota transplantation from Cd-exposed animals to control mice was performed, suggests that gut microbiota play a significant role in the entire process.

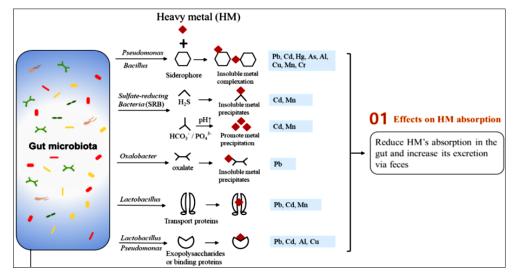


Fig 4: Direct effects of gut microbiota on the absorption of heavy metals (Duan et al., 2020) [11]

The interplay between Androgen and gut microbiota

The primary active forms of androgen, testosterone (T) and dihydrotestosterone (DHT), are the cornerstones for the development and maintenance of masculinization, normal sexual function, and spermatogenesis. Androgen has always served as a link between the endocrine system and the reproductive system (Matsumoto et al., 2013) [25]. Nearly 37.8% of senior men have testosterone insufficiency, also known as hypogonadism, which causes obesity, decreased glucose tolerance, metabolic syndrome, osteoporosis, etc (Mulligan et al., 2006) [26]. As the title suggests, multiple processes allow testosterone and gut microbiota to alter one another. Most of the bacteria are found in the gut, and it is known that the colon and rectum are where the androgen receptor is specifically expressed. Therefore, the gut microbes are constantly close to the androgen receptor. It is well established that androgen regulates stromal cell BMP and maintaining Wnt/TGFsignaling, intestinal homeostasis. As a result, androgen regulates the intestinal barrier and microenvironment, changing the gut flora. Additionally, it should be mentioned that the gut microbiota varies depending on gender. Prevotella is intimately correlated with testosterone synthesis, therefore men have higher levels of it. Bile aids in the direct excretion of glucuronidated androgens into the small intestine. Degluronidated testosterone (T-G) and dihydrotestosterone (DHT-G) are produced by the glucuronidase-dependent gut microbiota in the cecum before being absorbed in the distal intestine. Additionally, this testosterone production encourages the development of microbes. Additionally, the Clostridium and Ruminococcus species have a remarkable capacity to transform the androgens glucocorticoids, pregnenolone, and hydroxypregnenolone (J.M. et al., 2013).

In conclusion, it has been discovered that gut microbiota including Butyricoccus desmolans, Clostridium cadaveris, and Clostridium scindens express steroid processing enzymes like 17-Hydroxysteroid (17-HSD),3-HSD. As a result, levels of T metabolites, including DHT, and 3-diol concentrations, are found to be higher in the colon than in plasma (Devendran *et al.*, 2017) ^[6].

Several Fecal Microbiota Transplantation (FMT) studies have been conducted and showed that different gut microbiota can either enhance or disrupt the process of spermatogenesis. When FMT from High-fat diet (HFD) mice to normal diet mice were done, it disrupted the spermatogenesis process.

Fecal Microbiota Transplantation

To reconstruct gut dysbiosis, a treatment known as fecal microbiota transplantation involves administering a fecal matter solution that has been collected from a healthy donor animal into the recipient's intestinal tract. FMT involves several major phases, including choosing the donor animal, gathering the feces, processing them, preparing the recipient, and then transferring the feces to the recipient (Smits et al., 2013) [36]. There are several ways to administer feces including colonoscopy, nasogastric tube, and nasojejunal tube (Postigo et al., 2012) [30]. When it successfully treated Clostridium difficile illness, FMT gained notoriety (CDI). By inhibiting the pathogenic microbiota competitively, FMT restores the normal gut flora (Choi et al., 2016). It inhibits the pathogenic microbiota through the production of secondary bile acids from primary bile acids, boosting the production of free amino acids, and increasing the production of SCFA thereby lowering the pH. (Stellwag and Hylemon,1978) [38]. As has already been said in this book chapter, heavy metals modify the gut microbiota, and vice versa, the gut microbiota also has an impact on the absorption of heavy metals. The abundance of beneficial bacteria in the gut, including Bifidobacteria, Lactobacillus, Butyricoccus, etc., is decreased by heavy metal toxicity. Therefore, when feces from a healthy donor are given to the patient, the quantity of beneficial microbes quickly increases, ultimately improving the gut barrier, immunity, and SCFA synthesis. Additionally, these microbes will bind with heavy metal and produce toxic heavy metal precipitates, aiding in its elimination. Even yet, more investigation is needed to clarify the mechanism through which FMT reduces the toxicity of heavy metals.

Conclusion

Exposure to heavy metals has significantly increased as a result of excessive industrialization. Heavy metals are those metals that are hazardous to humans and have a higher density. A few examples of heavy metals are cadmium, lead, arsenic, and mercury. Because most herbicides, insecticides, batteries, automotive exhaust, etc. contain cadmium, which people are exposed to daily and eventually accumulate in the body for many years, cadmium has now become the element that poses the greatest risk to humans. Males are exposed to cadmium either through their environment or at work, which damages their reproductive system and causes asthenospermia, oligospermia, and azoospermia. It has been established that the blood-testis barrier is Cadmium's main target This is the case because BTB contains the cadmium-susceptible nectin protein and basal ES. Additionally, it inhibits the STAR gene, the Luteinizing Hormone Receptor (LHR), and the Androgen Receptor (AR), which are essential for the efficient operation of spermatogenesis. In this situation, the gut microbiota is important because heavy metals and gut bacteria interact. Metals can be chelated by microbiota like Lactobacillus and Pseudomonas, particularly Cd, which is then precipitated in the path. The system is protected by the gut microbiota in various ways, including by reducing pH, upping SCFA synthesis, boosting immunological function, and many others. Fecal microbiota transplantation has therefore emerged in the middle as the answer to lowering heavy metal toxicity, particularly cadmium. Fecal matter contains millions of gut microbes that, when properly fed into the colon, can outperform harmful bacteria in several ways, boosting the immune system and reinforcing the gut barrier. The FMT effect on heavy metal toxicity needs to be confirmed by more studies without a doubt.

References

- Arteaga Silva M, Arenas Rios E, Bonilla Jaime H, Damian Matzumura P, Limon Morales O, et al. Neuroendocrine effects of cadmium exposure on male reproductive functions. Frontiers in Bioscience-Landmark, 2020:26(2):286-326.
- 2. Bridou R, Monperrus M, Gonzalez PR, Guyoneaud R, Amouroux D. Simultaneous determination of mercury methylation and demethylation capacities of various sulfate-reducing bacteria using species-specific isotopic tracers. Environmental toxicology and chemistry,2011:30(2):337-344.
- 3. Chung NP, Cheng CY. Is cadmium chloride-induced inter-Sertoli tight junction permeability barrier disruption a suitable *in vitro* model to study the events

- of junction disassembly during spermatogenesis in the rat testis? Endocrinology,2001:142(5):1878-1888.
- 4. Chung NP, Cheng CY. Is cadmium chloride-induced inter-Sertoli tight junction permeability barrier disruption a suitable *in vitro* model to study the events of junction disassembly during spermatogenesis in the rat testis? Endocrinology,2001:142(5):1878-1888.
- 5. Clough SR, Welsh MJ, Payne AH, Brown CD, Brabec MJ. Primary rat Sertoli and interstitial cells exhibit a differential response to cadmium. Cell biology and toxicology,1990:6(1):63-79.
- Devendran S, Méndez García C, Ridlon JM. Identification and characterization of a 20β-HSDH from the anaerobic gut bacterium Butyricicoccus desmolans ATCC 43058 [S]. Journal of lipid research, 2017:58(5):916-925.
- 7. Dheer R, Patterson J, Dudash M, Stachler EN, Bibby KJ, Stolz DB, *et al.* Arsenic induces structural and compositional colonic microbiome change and promotes host nitrogen and amino acid metabolism. Toxicology and applied pharmacology,2015:289(3):397-408.
- 8. Diez Pina JM, Fernandez Aceñero MJ, Llorente Alonso MJ, Diaz Lobato S, Mayoralas S, Florez A. Tumor necrosis factor-alpha as a marker of systemic and local inflammation in "healthy" smokers. International journal of general medicine, 2009:2:9.
- 9. Ding N, Zhang X, Di Zhang X, Jing J, Liu SS, Mu YP, *et al.* Impairment of spermatogenesis and sperm motility by the high-fat diet-induced dysbiosis of gut microbes. *Gut*.2020:69(9):1608-1619.
- Dixon ZR, Shie FS, Warden BA, Burri BJ, Neidlinger TR. The effect of a low carotenoid diet on malondialdehyde-thiobarbituric acid (MDA-TBA) concentrations in women: a placebo-controlled doubleblind study. Journal of the American College of Nutrition, 1998:17(1):54-58.
- 11. Duan H, Yu L, Tian F, Zhai Q, Fan L, Chen W. Gut microbiota: A target for heavy metal toxicity and a probiotic protective strategy. Science of the Total Environment,2020:742:140429.
- 12. Elgawish RAR, Ghanem ME. Effect of long-term cadmium chloride exposure on testicular functions in male albino rats. American Journal of Animal and Veterinary Sciences, 2014:9(4):182.
- 13. Faroon O, Ashizawa A, Wright S, Tucker P, Jenkins K, Ingerman L, *et al.* Toxicological profile for cadmium, 2013.
- 14. Fazeli M, Hassanzadeh P, Alaei S. Cadmium chloride exhibits a profound toxic effect on bacterial microflora of the mice gastrointestinal tract. Human & experimental toxicology,2011:30(2):152-159.
- 15. Fels J, Scharner B, Zarbock R, Zavala Guevara IP, Lee WK, Barbier OC, *et al.* Cadmium complexed with β2-microglobulin, albumin, and lipocalin-2 rather than metallothionein cause megalin: cubilin dependent toxicity of the renal proximal tubule. International Journal of Molecular Sciences, 2019:20(10):2379.
- 16. Fujishiro H, Hamao S, Tanaka R, Kambe T, Himeno S. Concentration-dependent roles of DMT1 and ZIP14 in cadmium absorption in Caco-2 cells. The Journal of Toxicological Sciences, 2017:42(5):559-567.
- 17. Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal

- bacteriotherapy) for recurrent Clostridium difficile infection. Clinical infectious diseases,2011:53(10):994-1002.
- 18. Henson MC, Chedrese PJ. Endocrine disruption by cadmium, a common environmental toxicant with paradoxical effects on reproduction. Experimental biology and medicine,2004:229(5):383-392.
- 19. Human Microbiome Jumpstart Reference Strains Consortium, Nelson KE, Weinstock GM, Highlander SK, Worley KC, Creasy HH, Wortman JR, *et al.* A catalog of reference genomes from the human microbiome. Science, 2010:328(5981):994-999.
- 20. Janecki A, Jakubowiak A, Steinberger A. Effect of cadmium chloride on the transepithelial electrical resistance of Sertoli cell monolayers in two-compartment cultures—a new model for toxicological investigations of the "blood-testis" barrier *in vitro*. Toxicology and applied pharmacology,1992:112(1):51-57.
- 21. Kidd SA, Eskenazi B, Wyrobek AJ. Effects of male age on semen quality and fertility: a review of the literature. Fertility and sterility, 2001:75(2):237-248.
- 22. Kilchevsky A, Honig S. Male factor infertility in 2011: semen quality, sperm selection, and 494 hematospermia. *Nat Rev Urol*, 2012:9:68-70.
- 23. Lafuente A, Cano P, Esquifino AI. Are cadmium effects on plasma gonadotropins, prolactin, ACTH, GH and TSH levels, dose-dependent?. *Biometals*, 2003:16(2):243-250.
- 24. Li X, Cheng W, Shang H, Wei H, Deng C. The interplay between androgen and gut microbiota: is there a microbiota-gut-testis Axis? Reproductive Sciences, 2021, 1-11.
- 25. Matsumoto T, Sakari M, Okada M, Yokoyama A, Takahashi S, Kouzmenko A, *et al.* The androgen receptor in health and disease. *Annu Rev Physiol*,2013:75(1):201-224.
- 26. Mulligan T, Frick MF, Zuraw QC, Stemhagen A, McWhirter C. Prevalence of hypogonadism in males aged at least 45 years: the HIM study. International journal of clinical practice, 2006:60(7):762-769.
- 27. Oliva R. Protamines and male infertility. Human reproduction update, 2006:12(4):417-435.
- 28. Pant N, Banerjee AK, Pandey S, Mathur N, Saxena DK, Srivastava SP. Correlation of lead and cadmium in human seminal plasma with seminal vesicle and prostatic markers. Human & experimental toxicology, 2003:22(3):125-128.
- 29. Papanicolas LE, Choo JM, Wang Y, Leong LE, Costello SP, Gordon DL, *et al.* Bacterial viability in faecal transplants: which bacteria survive?. *EBioMedicine*, 2019:41:509-516.
- 30. Postigo R, Kim JH. Colonoscopic versus nasogastric fecal transplantation for the treatment of Clostridium difficile infection: a review and pooled analysis. Infection, 2012:40(6):643-648.
- 31. Prozialeck WC, Lamar PC, Lynch SM. Cadmium alters the localization of N-cadherin, E-cadherin, and β-catenin in the proximal tubule epithelium. Toxicology and applied pharmacology,2003:189(3):180-195.
- 32. Ridlon JM, Ikegawa S, Alves JM, Zhou B, Kobayashi A, Iida T, *et al.* Clostridium scindens: a human gut microbe with a high potential to convert glucocorticoids

- into androgens. Journal of lipid research, 2013:54(9):2437-2449.
- 33. Robaire B, Hinton BT. The Epididymis. In 'Knobil and Neill's Physiology of Reproduction, 4th edn, 2015.
- 34. Siu ER, Mruk DD, Porto CS, Cheng CY. Cadmium-induced testicular injury. Toxicology and applied pharmacology,2009:238(3):240-249.
- 35. Siu MK, Wong CH, Lee WM, Cheng CY. Sertoli-germ cell anchoring junction dynamics in the testis are regulated by an interplay of lipid and protein kinases. Journal of Biological Chemistry, 2005:280(26):25029-25047.
- Smits LP, Bouter KE, De Vos WM, Borody TJ, Nieuwdorp M. Therapeutic potential of fecal microbiota transplantation. Gastroenterology, 2013:145(5):946-953.
- 37. Sorenson DR, Brabec MJ. The Responce of Adult Rat Sertoli Cells, Immortalized by a Temperature-sensitive mutant of SV40, to 1, 2-dinitrobenzene, 1, 3-dinitrobenzene, 2, 4-dinitrotoluene, 3, 4-dinitrotoluene, and cadmium, 2003.
- 38. Stellwag EJ, Hylemon PB. Characterization of 7-α-dehydroxylase in Clostridium leptum. The American journal of clinical nutrition,1978:31(10):S243-S247.
- 39. SYED V, GU W, HECHT NB. Sertoli cells in culture and mRNA differential display provide a sensitive early warning assay system to detect changes induced by xenobiotics. Journal of andrology,1997:18(3):264-273.
- 40. Takai Y, Nakanishi H. Nectin and afadin: novel organizers of intercellular junctions. Journal of cell science, 2003:116(1):17-27.
- 41. Tardif S, Dubé C, Chevalier S, Bailey JL. Capacitation is associated with tyrosine phosphorylation and tyrosine kinase-like activity of pig sperm proteins. Biology of reproduction, 2001:65(3):784-792.
- 42. Thévenod F, Lee WK. Cadmium and cellular signaling cascades: interactions between cell death and survival pathways. Archives of toxicology,2013:87(10):1743-1786.
- 43. Wan HT, Mruk DD, Wong CK, Cheng CY. The apical ES-BTB-BM functional axis is an emerging target for toxicant-induced infertility. *Trends Mol Med*, 2013:19:396-405.
- 44. Wang L, Li Y, Fu J, Zhen L, Zhao N, Yang Q, *et al.* Cadmium inhibits mouse sperm motility through inducing tyrosine phosphorylation in a specific subset of proteins. Reproductive toxicology,2016:63:96-106.
- 45. Wang Z, Sun Y, Yao W, Ba Q, Wang H. Effects of cadmium exposure on the immune system and immunoregulation. Frontiers in Immunology, 2021, 2892.
- 46. WHO (World Health Organization): Exposure to cadmium: a major public health concern, Public Health, and Environment WHO. 20 Avenue Appia, 1211 Geneva 27, Switzerland, 2010.
- 47. Wong CH, Mruk DD, Lui WY, Cheng CY. Regulation of blood-testis barrier dynamics: an *in vivo* study. Journal of cell science,2004:117(5):783-798.
- 48. Yan HH, Cheng CY. Blood–testis barrier dynamics are regulated by an engagement/disengagement mechanism between tight and adherens junctions via peripheral adaptors. Proceedings of the National Academy of Sciences,2005:102(33):11722-11727.

- 49. Yu X, Hong S, Faustman EM. Cadmium-induced activation of stress signaling pathways, disruption of ubiquitin-dependent protein degradation and apoptosis in primary rat Sertoli cell-gonocyte cocultures. Toxicological sciences, 2008:104(2):385-396.
- 50. Zhou D, Pan Q, Shen F, Cao HX, Ding WJ, Chen YW, *et al.* Total fecal microbiota transplantation alleviates high-fat diet-induced steatohepatitis in mice via beneficial regulation of gut microbiota. Scientific reports, 2017:7(1):1-11.