



LPS triggers liver TLR4 signaling from the gastrointestinal tract during SARA

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Abstract

Intake of huge-concentration of diet for an expanded period can cause of SARA. Subacute ruminal acidosis also a severe cause of the decline in milk quality and quantity. The Subacute ruminal acidosis (SARA) has identified to stimulate a systemic inflammatory response. That is possibly caused by the dislocation of lipopolysaccharides (LPS) from the digestive tract to the circulatory system.

Keywords: subacute ruminal acidosis, inflammatory response, lipopolysaccharides

Introduction

Subacute ruminal acidosis (SARA) is a prevalent digestive disorder [1, 2]. The outcome of SARA has contained depression of food intake, diarrhea, decreased ruminal pH, laminitis, and inflammatory response. Which unseasonable affected the milk quality and quantity result in the depression [3, 7]. This decrease in pH through extreme high concentrate (HC) diet the unremitting using of HC diet by ruminants it caused to enhances the triggered of the pH and a depression production of the organic acids in the digestive organs [8, 10]. Together, these changes can finally lead to the congregation of lipopolysaccharides (LPS). Also, it can simultaneously disrupt the gastrointestinal barrier and assist the translocation of LPS from the digestive tract into the circulatory system, often induction a systemic inflammatory response [11, 12]. It has significantly recognized that HC-induced SARA intensify the plasma concentrations of acute-phase proteins (APPs), like as haptoglobin (Hp), serum amyloid A (SAA), and LPS-binding protein (LBP), and raise the rats of pro-inflammatory cytokines, including TNF- α , and interleukin (IL)-1, IL-6 in the peripheral blood and induce APP synthesis via their special hepatic receptors [6, 13, 15]. TLR4 identified LPS with the help of the LBP and cluster of CD14 [16, 17]. MyD88 has activated by LPS after connecting with TLR4 on the surface of the host cell [18]. The hepatic organ is the principal and first place for the synthesis of APPs, including LBP, SAA, C-reactive protein (CRP), and HP [19]. These immune response genes contain the cytokine encoding genes whose expressions have intensely enhanced in the livers of cows as a consequence of LPS-induced mastitis [20, 21]. However, it is unrecognized whether blood circulating obtained LPS from the digestive tract of ruminant. Then can increase the expressions of liver immune response genes through the TLR4-NF- κ B signaling pathway [22, 23] (see Figure) [24]. Accordingly, we hypothesized that the enhanced displacement of lipopolysaccharide (LPS) *via* the portal vein from the digestive tract into the liver, conceivably result in the epigenetically modulated expression of TLR4 and consequently activated the TLR4-NF- κ B pathway and

finally trigger the enhanced expression of immune response genes in this organ.

SARA ruminants represented substantial enhanced LPS concentrations in portal vein and rumen. The hepatics of these ruminants depicted increased mRNA concentrations of proinflammatory genes that demonstrated inflammation. However, the occurrence of hepatic inflammation has further accredited by the increased protein expression of those cytokines in the hepatics of SARA. These intensified expressions of well-known pro-inflammatory genes were seemingly interceded by compelled TLR4 signaling because SARA raised the concentrations of protein and TLR4 mRNA in hepatic and the plenty of the NF- κ B-p65 factor and its active phosphorylated varied.

A significant adverse feature of diet-induced SARA is causing liver inflammation. This inflammation has triggered by enhanced rates of circulating LPS that, in turn, possibly activate extreme TLR4 signaling, the prevalent consequences of liver dysfunction may also affect milk production [8].

Conflict of Interest Statement

The authors declare there are no conflicts of interest

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References

1. Bilal MS, Abaker JA, ul Aabdin Z, Xu T, Dai H, Zhang K, *et al.* Lipopolysaccharide derived from the digestive tract triggers an inflammatory response in the uterus of mid-lactating dairy cows during SARA. BMC Vet Res [Internet]. 2016; 12(1):1-7. Available from: <http://dx.doi.org/10.1186/s12917-016-0907-1>
2. Steele MA, Croom J, Kahler M, Alzahal O, Hook SE, Plaizier K, *et al.* Bovine rumen epithelium undergoes rapid structural adaptations during grain-induced subacute ruminal acidosis. Am J Physiol - Regul Integr Comp Physiol. 2011; 300(6):1515-23.

3. Kleen JL, Hooijer GA, Rehage J, Noordhuizen JPTM. Subacute ruminal acidosis (SARA): A review. *J Vet Med Ser A Physiol Pathol Clin Med.* 2003; 50(8):406-14.
4. Plaizier JC, Krause DO, Gozho GN, McBride BW. Subacute ruminal acidosis in dairy cows: The physiological causes, incidence and consequences. *Vet J [Internet].* 2008; 176(1):21-31. Available from: <http://dx.doi.org/10.1016/j.tvjl.2007.12.016>
5. Gozho GN, Plaizier JC, Krause DO, Kennedy AD, Wittenberg KM. Subacute ruminal acidosis induces ruminal lipopolysaccharide endotoxin release and triggers an inflammatory response. *J Dairy Sci [Internet].* 2005; 88(4):1399-403. Available from: [http://dx.doi.org/10.3168/jds.S0022-0302\(05\)72807-1](http://dx.doi.org/10.3168/jds.S0022-0302(05)72807-1)
6. Dong H, Wang S, Jia Y, Ni Y, Zhang Y, Zhuang S. *et al.* Long-term effects of Subacute Ruminal Acidosis (SARA) on milk quality and hepatic gene expression in lactating goats fed a high-concentrate diet. *PLoS One,* 2013; 8(12).
7. Colman E, Fokink WB, Craninx M, Newbold JR, De Baets B, Fievez V, *et al.* Effect of induction of subacute ruminal acidosis on milk fat profile and rumen parameters. *J Dairy Sci [Internet].* 2010; 93(10):4759-73. Available from: <http://dx.doi.org/10.3168/jds.2010-3158>
8. Chang G, Zhuang S, Seyfert HM, Zhang K, Xu T, Jin D. *et al.* Hepatic TLR4 signaling is activated by LPS from digestive tract during SARA, and epigenetic mechanisms contribute to enforced TLR4 expression. *Oncotarget.* 2015; 6(36):38578-90.
9. Li S, Khafipour E, Krause DO, Kroeker A, Rodriguez-Lecompte JC, Gozho GN. *et al.* Effects of subacute ruminal acidosis challenges on fermentation and endotoxins in the rumen and hindgut of dairy cows. *J Dairy Sci.* 2012; 95(1):294-303.
10. Garrett EF, Pereira MN, Nordlund KV, Armentano LE, Goodger WJ, Oetzel GR. *et al.* Diagnostic methods for the detection of subacute ruminal acidosis in dairy cows. *J Dairy Sci [Internet].* 1999; 82(6):1170-8. Available from: [http://dx.doi.org/10.3168/jds.S0022-0302\(99\)75340-3](http://dx.doi.org/10.3168/jds.S0022-0302(99)75340-3)
11. Liu J hua, Xu T ting, Zhu W yun, Mao S yong. A high-grain diet alters the omasal epithelial structure and expression of tight junction proteins in a goat model. *Vet J [Internet].* 2014; 201(1):95-100. Available from: <http://dx.doi.org/10.1016/j.tvjl.2014.03.025>
12. Khafipour E, Krause DO, Plaizier JC. A grain-based subacute ruminal acidosis challenge causes translocation of lipopolysaccharide and triggers inflammation. *J Dairy Sci [Internet].* 2009; 92(3):1060-70. Available from: <http://dx.doi.org/10.3168/jds.2008-1389>
13. Jia YY, Wang SQ, Ni YD, Zhang YS, Zhuang S, Shen XZ. High concentrate-induced subacute ruminal acidosis (SARA) increases plasma acute phase proteins (APPs) and cortisol in goats. *Animal.* 2014; 8(9):1433-8.
14. Zhou J, Dong G, Ao C, Zhang S, Qiu M, Wang X. *et al.* Feeding a high-concentrate corn straw diet increased the release of endotoxin in the rumen and pro-inflammatory cytokines in the mammary gland of dairy cows. *BMC Vet Res.* 2014; 10(1):1-10.
15. Heinrich PC, Behrmann I, Müller-Newen G, Schaper F, Graeve L. Interleukin-6-type cytokine signalling through the gp130/Jak/STAT pathway. *Biochem J.* 1998; 334(2):297-314.
16. Sohn MJ, Hur GM, Byun HS, Kim WG. Cyclo (dehydrohistidyl-L-tryptophyl) inhibits nitric oxide production by preventing the dimerization of inducible nitric oxide synthase. *Biochem Pharmacol.* 2008; 75(4):923-30.
17. Bannerman DD, Paape MJ, Hare WR, Hope JC. Characterization of the bovine innate immune response to intramammary infection with *Klebsiella pneumoniae*. *J Dairy Sci [Internet].* 2004; 87(8):2420-32. Available from: [http://dx.doi.org/10.3168/jds.S0022-0302\(04\)73365-2](http://dx.doi.org/10.3168/jds.S0022-0302(04)73365-2)
18. Ju J, Li L, Xie J, Wu Y, Wu X, Li W. *et al.* Toll-like receptor-4 pathway is required for the pathogenesis of human chronic endometritis. *Exp Ther Med.* 2014; 8(6):1896-900.
19. Murata H, Shimada N, Yoshioka M. Current research on acute phase proteins in veterinary diagnosis: An overview. *Vet J.* 2004; 168(1):28-40.
20. Jiang L, Sørensen P, Røntved C, Vels L, Ingvarsen KL. Gene expression profiling of liver from dairy cows treated intra-mammary with lipopolysaccharide. *BMC Genomics.* 2008; 9:1-12.
21. Vels L, Røntved CM, Bjerring M, Ingvarsen KL. Cytokine and acute phase protein gene expression in repeated liver biopsies of dairy cows with a lipopolysaccharide-induced mastitis. *J Dairy Sci [Internet].* 2009; 92(3):922-34. Available from: <http://dx.doi.org/10.3168/jds.2008-1209>
22. Liu S, Shi X, Bauer I, Günther J, Seyfert HM. Lingual antimicrobial peptide and IL-8 expression are oppositely regulated by the antagonistic effects of NF- κ B p65 and C/EBP β in mammary epithelial cells. *Mol Immunol [Internet].* 2011; 48(6-7):895-908. Available from: <http://dx.doi.org/10.1016/j.molimm.2010.12.018>
23. Wiench M, Miranda TB, Hager GL. Control of nuclear receptor function by local chromatin structure. *FEBS J.* 2011; 278(13):2211-30.
24. Ceccarelli S, Nobili V, Alisi A. Toll-like receptor-mediated signaling cascade as a regulator of the inflammation network during alcoholic liver disease. *World J Gastroenterol.* 2014; 20(44):16443-51.